

**HAEMATOLOGICAL EFFECTS OF THE AQUEOUS EXTRACT OF *Acanthus*
montanus LEAF IN MALE WISTAR RATS**

BY

Blessing Eloho EREKWEDOR

LSC2007295

DEPARTMENT OF SCIENCE LABORATORY TECHNOLOGY

(BIOTECHNOLOGY TECHNIQUES)

FACULTY OF LIFE SCIENCE

UNIVERSITY OF BENIN

BENIN CITY.

OCTOBER, 2025.

HAEMATOLOGICAL EFFECTS OF THE AQUEOUS EXTRACT OF *Acanthus montanus* LEAF IN MALE WISTAR RATS

BY

Blessing Eloho EREWEDOR

LSC2007295

**A PROJECT WORK SUBMITTED TO THE DEPARTMENT OF SCIENCE
LABORATORY TECHNOLOGY, FACULTY OF LIFE SCIENCES, UNIVERSITY OF
BENIN, BENIN CITY, IN PARTIAL FULFILMENT OF THE REQUIREMENTS FOR
THE AWARD OF BACHELORS OF SCIENCE DEGREE (BSc.) IN SCIENCE
LABORATORY TECHNOLOGY (BIOTECHNOLOGY TECHNIQUES)**

OCTOBER, 2025.

AUTHOR'S STATEMENT

I hereby grant the University of Benin, through the University of Benin Library, a non-exclusive, worldwide right to reproduce and distribute my thesis and abstract (hereinafter "the Work"), in whole or in part, through any media, in its present form or any translated version for preservation and accessibility, provided such translation does not alter its content. This grant is royalty-free, and I retain the right to publish the Work in its current or future versions elsewhere.

Warranties

I further affirm that:

1. I am the sole author of the Work and grant the University of Benin the right to make it available five (5) years after the award of my degree, in compliance with the University of Benin Senate regulations.
2. The Work does not contain confidential information requiring third-party consent for disclosure.
3. I have exercised due diligence to ensure that the Work is original and does not breach any Nigerian law or infringe upon any third party's copyright or other Intellectual Property Rights, to the best of my knowledge.
4. Where the Work includes copyrighted material not owned by me, I have obtained unrestricted permission from the copyright holder to grant this license to the University of Benin Library. Such third-party materials are clearly identified and acknowledged within the Work.
5. In the event of any copyright dispute concerning the Work, I agree to indemnify and hold harmless the University of Benin, its officers, employees, and agents from any liability arising from the material authorized under this agreement.
6. The University of Benin is under no obligation whatsoever to take legal action on my behalf as the Depositor in the event of an intellectual property rights infringement or any other related dispute in the material deposited.

Author's Name

Signature/Date

Email

Supervisor's Name

Signature/Date

Email

Supervisor's Name

Signature/Date

Email

CERTIFICATION

This is to certify that this project work titled "**Haematological effects of the aqueous extract of *Acanthus montanus* leaf in male Wistar rats**" was completed by **Blessing Eloho EREKWEDOR (Miss)** with matriculation number **LSC2007295**, Department of Science Laboratory Technology (Biotechnology Techniques), Faculty of Life Sciences, University of Benin, Benin City.

Dr Joseph O. Erhabor
(Project Supervisor)

Date

Dr. P. O. ALONGE
(Project Co-ordinator)

Date

Prof. J. O. OSARUMWENSE
(Head of Department)

Date

(EXTERNAL EXAMINER).

Date

DECLARATION

I declare that the project work titled- Haematological effects of the aqueous extract of *Acanthus montanus* leaf in male Wistar rats” was written by me in the Department of Science Laboratory Technology (Biotechnology Techniques), University of Benin, Benin City, Edo State.

Blessing Eloho EREKWEDOR (Miss).
(Student)

DATE

DEDICATION

I declare that the project work titled- Haematological effects of the aqueous extract of *Acanthus montanus* leaf in male Wistar rats” was written by me in the Department of Science Laboratory Technology (Biotechnology Techniques), University of Benin, Benin City, Edo State.

ACKNOWLEDGEMENTS

First, my sincere gratitude goes to God Almighty for his constant guidance, provision, unwavering love and protection through my stay at the University of Benin.

I extend my sincere gratitude and acknowledgement to my project supervisor, Dr J.O. Erhabor for his time, effort, and ever-receptive ears towards me, which guided me every step of the way through the completion of this project.

My special thanks go to my dear Parent, Mr. and Mrs. EREKWEDOR, for their unfailing love, words of encouragement, and support.

TABLE OF CONTENTS

Cover Page	i
Title Page	ii
Certification	iv
Declaration	iv
Dedication	vi
Acknowledgement	vii
Table of contents	viii
List of Tables	xi
CHAPTER ONE	1
1.0 INTRODUCTION	1
1.1 Background of the Study	1
1.2 A Brief Overview of Haematology	2
1.3 Botanical Description of <i>Acanthus montanus</i>	2
1.4 Folklore Uses of <i>Acanthus montanus</i>	2
1.5 Reported Pharmacological Uses of <i>Acanthus montanus</i>	3
1.6 Fertility and Haematological Study	4

1.7 Aim of the Study	4
1.8 Objectives of the Study	5
CHAPTER TWO	6
2.0 LITERATURE REVIEW	6
2.1 Phytochemical Composition of <i>A.motanus</i>	6
2.2 Haematological Effects	7
2.3 Clinical Applications and Potential Health Benefits	7
2.4 Toxic Properties	8
2.4.1 Subacute Toxicity Assessment:	9
CHAPTER THREE	10
3.0 MATERIALS AND METHODS	10
3.1 Materials Used	10
3.1 Collection of the Plant sample	10
3.2 Preparation and Extraction	10
3.3 Experimental Animal	11
3.4 Experimental Design	11
3.6 Data Analysis	11

CHAPTER FOUR	12
4.0 RESULTS	12
4.1 Effect of <i>Acanthus motanus</i> leaf on white blood cells and their differentials in rats.	12
4.2: Effect of aqueous extract of <i>Acanthus motanus</i> on red blood cell and its differential in Male Wistar rats	13
4.3: Effect of 14-day administration of aqueous extract of <i>Acanthus motanus</i> on Platelets and their Factors in Wistar rats	14
CHAPTER FIVE	15
5.0 DISCUSSION	15
CONCLUSION	17
REFERENCES	18

LIST OF TABLES

TABLE 4.1: White blood cell and differentials count in Wistar rats subjected to 14 days administration of aqueous extract of <i>Acanthus motanus</i> .	12
TABLE 4.2: Red blood cell and differentials count in Wistar rats subjected to 14 days oral administration of aqueous extract of <i>Acanthus motanus</i> .	13
TABLE 4.3: Effect of 14-day administration of aqueous extract of <i>Acanthus motanus</i> on Platelets and their Factors in Wistar rats	14

ABSTRACT

The study investigated the effect of *Acanthus montanus* leaf's aqueous extract on male Wistar rats' haematological parameters. The research aimed to evaluate the ability of the plant extract to protect and restore normal haematological parameters following administration of the extract. Fifteen male Wistar rats were divided into control and experimental groups. The experimental groups received varying doses of the aqueous extract of *Acanthus montanus* orally for a specified period. Blood samples were analysed to determine haematological indices, including packed cell volume (PCV), haemoglobin concentration (HB), red blood cell count (RBC), white blood cell count (WBC), and differential leukocyte count. Results revealed that administration of *Acanthus montanus* extract significantly ($p < 0.05$) improved haematological parameters in a dose-dependent manner compared to the untreated group. The extract normalised the levels of packed cell volume (PCV), haemoglobin (Hb), and red blood cell (RBC) count, while also stabilising white blood cell (WBC) and lymphocyte counts. The study indicates that *Acanthus montanus* possesses potent haematoprotective activity, likely due to its phytoconstituents such as flavonoids, tannins, alkaloids, and saponins, which help mitigate oxidative damage to blood cells. Therefore, the aqueous leaf extract of *Acanthus montanus* may serve as a potential natural therapeutic agent for managing haematological disorders and conditions associated with blood toxicity. Further studies are recommended to isolate and characterise the active compounds responsible for its protective effects.

CHAPTER ONE

1.0

INTRODUCTION

1.1 Background of the Study

The increasing global interest in medicinal plants has led to intensive research to validate traditional claims and discover potential therapeutic agents. In particular, the search for haematoprotective compounds from natural sources is gaining traction, especially in drug-induced haematological toxicity, environmental stress, and disease-related blood disorders (Odey *et al.*, 2012). Haematological parameters are vital in assessing the physiological and pathological conditions of organisms and serve as indicators of bone marrow activity, immune response, and the general well-being of animals and humans

(Mukinda and Syce, 2007). *Acanthus montanus*, commonly known as Mountain Thistle or Bear's Breech, is widely used in traditional African medicine to treat various ailments such as wounds, inflammation, and infections. However, limited scientific studies have validated its use in maintaining or restoring haematological health. Investigating the haematoprotective effect of the aqueous extract of *Acanthus montanus* could provide empirical support for its traditional applications and open pathways for developing novel natural therapies for blood-related conditions. This study is designed to assess the potential protective effects of *Acanthus montanus* extract on the blood profile of male Wistar rats and determine its influence on selected haematological parameters.

1.2 A Brief Overview of Haematology

Haematology is the branch of biomedical science that deals with studying blood, blood-forming organs, and blood diseases. It encompasses the analysis of cellular components such as red blood cells (RBCs), white blood cells (WBCs), platelets, haemoglobin levels, haematocrit, and related indices (Guyton and Hall, 2011). These parameters are essential for diagnosing conditions like anaemia, infections, and blood-clotting disorders. In experimental pharmacology, haematological parameters are routinely used to assess the toxicological or protective effects of plant extracts and drugs. Alterations in these values may signify adverse or beneficial effects on the bone marrow or immune system (Yakubu *et al.*, 2007). Thus, this study evaluates how *Acanthus montanus* extract may affect hematopoietic function.

1.3 Botanical Description of *Acanthus montanus*

Acanthus montanus (Nees) T. Anderson belongs to the family Acanthaceae. It is a fast-growing, evergreen, perennial shrub native to tropical regions of Africa. The plant is characterized by its spiny-margined leaves, purplish tubular flowers, and erect woody stems that may reach heights up to 2 meters. The leaves are opposite, ovate, dark green, and shiny, and the flowers are borne in spikes (Burkill, 1985). It thrives in moist, shaded environments such as forest under stories and riverbanks. The plant is often propagated via stem cuttings, and the parts commonly used in traditional medicine include the leaves, roots, and bark.

1.4 Folklore Uses of *Acanthus montanus*

In African traditional medicine, *Acanthus montanus* is employed for various ethnomedical purposes. Local communities use aqueous or alcoholic extracts of the plant for treating:

- Rheumatism and arthritis
- Gastrointestinal problems
- Wounds and sores
- Snakebites
- Respiratory tract infections
- Female infertility and dysmenorrhea

The leaves are often crushed and applied topically for wound healing, while decoctions are taken orally to treat internal conditions (Iwu, 1993). Despite its wide use in folk medicine, few pharmacological studies have validated these claims, thus warranting further scientific investigation.

1.5 Reported Pharmacological Uses of *Acanthus montanus*

Modern pharmacological investigations into *Acanthus montanus* have revealed several biological activities. These include:

Anti-inflammatory: Extracts of the plant have shown inhibition of inflammation in animals models (Mbah *et al.*, 2007).

Antioxidant: Phytochemical screening has demonstrated significant antioxidant potential due to flavonoid and phenolic contents (Ezekwesili *et al.*, 2014).

Wound Healing: Topical application accelerates wound contraction and re-epithelialization (Ezekwesili *et al.*, 2010).

Antimicrobial: Exhibits broad-spectrum activity against bacteria and fungi (El-Mahmood *et al.*, 2010).

Hepatoprotective: Some studies suggest protection of liver tissue from toxic injury (Chukwuma *et al.*, 2014).

These findings suggest that the plant may have therapeutic applications beyond traditional usage, but limited data exists regarding its haematological effects.

1.6 Fertility and Haematological Study

Although *Acanthus montanus* is traditionally used to treat reproductive issues, there is limited direct scientific evidence supporting its effects on male fertility. Some ethnobotanical sources suggest it may influence hormonal regulation or improve sexual function, but clinical studies remain sparse (Obianime and Roberts, 2009). Fertility and haematology are closely interlinked, as healthy blood parameters such as adequate hemoglobin levels and leukocyte counts play vital roles in hormonal transport, immune regulation, and tissue oxygenation. Thus, while this study focuses on haematological outcomes, the results may indirectly provide insight into the plant's effect on reproductive health.

1.7 Aim of the Study

This study aimed to investigate the effect of the aqueous extract of *Acanthus montanus* on haematological parameters in rats.

1.8 Objectives of the Study

The Objectives of the study were;

- i. To determine the effect of aqueous extract of *Acanthus montanus* on red blood cell (RBC), white blood cell (WBC), platelet count, and haemoglobin levels in male Wistar rats.
- ii. To assess the dose-dependent response of haematological parameters to varying concentrations of the plant extract.

CHAPTER TWO

2.0

LITERATURE REVIEW

Acanthus montanus (Nees) T. Anderson is a perennial shrub belonging to the family Acanthaceae. It is widely distributed in tropical Africa and used traditionally for treating inflammatory disorders, fever, pain, and infections. Recent pharmacological studies have reported antioxidant, anti-inflammatory, hepatoprotective, and haematological effects of its extracts. Since hepatotoxicity often involves oxidative stress and blood alterations, it is important to review the existing literature on the plant's phytochemical composition, haematological effects, clinical applications, and toxicological profile, especially subacute toxicity, to justify its scientific evaluation in hepatoprotection.

2.1 Phytochemical Composition of *A. motanus*

Phytochemicals are secondary metabolites responsible for medicinal plants' biological and therapeutic activities. Studies on *A. montanus* leaves, stems, and roots reveal a wide spectrum of compounds. Qualitative screenings report the presence of alkaloids, flavonoids, saponins, tannins, terpenoids, steroids, glycosides, phenols, and anthocyanins (Igwe *et al.*, 2017). Proximate composition of the leaves shows appreciable nutrients: carbohydrate (~37.86%), protein (~17.72%), fibre (~16.70%), ash (~10.56%), fat (~5.31%), and minerals such as calcium, magnesium, potassium, sodium, iron, zinc, and copper (Añuli *et al.*, 2020). Processing effects: boiling, sun-drying, and oven-drying significantly reduce levels of alkaloids, tannins, saponins, phenols, and anthocyanins, suggesting the raw plant may contain higher concentrations of active compounds (Igwe *et al.*, 2017). Isolated compounds: A new phenylethanoid glycoside named

acanmontanoside, along with known compounds such as verbascoside, isoverbascoside, and leucosceptoside A, have been identified, many of which are associated with antioxidant and hepatoprotective activity (Nguelefack *et al.*, 2010).

These findings suggest that *A. montanus* is rich in phytochemicals with antioxidant, anti-inflammatory, and membrane-stabilizing potential, which may underpin hepatoprotective effects.

2.2 Haematological Effects

Haematological indices are important markers of physiological and pathological conditions. Several studies show that *A. montanus* extracts exert beneficial effects on blood parameters, especially under oxidative or toxic stress (Uroko *et al.*, 2020). It also reported that the methanol extract of *A. montanus* leaves significantly improved RBC count, haemoglobin, PCV, and WBC count in acetaminophen-induced oxidative stress in rats. The extract also restored antioxidant enzymes: Superoxide dismutase (SOD), Catalase (CAT), Glutathione peroxidase (GPx) and reduced lipid peroxidation (MDA levels).

A hypolipidemic and haematological study (Osigwe *et al.*, 2024) found dose-dependent increases in haemoglobin and RBC count in rats treated with leaf extracts, suggesting erythropoietic stimulation.

2.3 Clinical Applications and Potential Health Benefits

Traditional medicine has long employed *A. montanus*, and scientific studies have validated several of its uses: Analgesic and anti-inflammatory: Methanol extracts significantly reduced pain and paw oedema in mice, confirming its folkloric use against pain and inflammation (Asongalem *et al.*, 2004).

Antidiabetic: Ethanol root extracts reduced blood glucose levels in alloxan-induced diabetic rats, indicating hypoglycemic activity (Odoh and Ezugwu, 2011).

Anthelmintic: Aqueous leaf extracts inhibited egg hatching and larval development of nematodes, supporting traditional use against helminth infections (Hounzangbé-Adoté *et al.*, 2010).

Anti-ulcer: Crude and fractionated extracts reduced gastric lesions in indomethacin-induced ulcer models (Nworu *et al.*, 2020).

Antimicrobial/Immunomodulatory: Root aqueous extract enhanced phagocytic activity, neutrophil count, and reduced microbial growth in experimental models (Nguelefack *et al.*, 2010).

Hemostatic: Methanol extract shortened bleeding and clotting time in rats, linked to flavonoids like rutin and catechin (Osigwe *et al.*, 2023).

Overall, the plant exhibits multi-target pharmacological properties that justify its continued evaluation for hepatoprotective activity.

2.4 Toxic Properties

Although generally regarded as safe in traditional use, experimental studies have identified potential toxicities.

Acute toxicity: Oral LD₅₀ values are > 5000 mg/kg in rodents, indicating wide safety margins (Osigwe *et al.*, 2023).

Reproductive toxicity: Ethanol leaf extract caused decreased sperm motility, viability, and spermicide count in male Wistar rats, suggesting possible antifertility effects (Anyanwu *et al.*, 2021).

Maternal/fetal toxicity: Pregnant rats treated with methanol/methylene chloride extracts (250–1000 mg/kg) showed reduced foetal weight and ossification, although neonates recovered postnatally (Asongalem *et al.*, 2008).

2.4.1 Subacute Toxicity Assessment:

Subacute studies show no mortality or overt toxicity when extracts are given daily for 14–28 days, but some biochemical alterations have been noted. Uroko *et al.* (2020) reported no adverse effects at hepatoprotective doses (200–500 mg/kg) for 14 days. Osigwe *et al.* (2023) found that repeated administration of methanol extract (200–800 mg/kg for 9 days) caused no behavioural toxicity. However, reproductive and embryotoxic findings highlight the need for careful dose standardization and safety assessment, especially for prolonged use.

CHAPTER THREE

3.0 MATERIALS AND METHODS

3.1 Materials Used

Materials used are a water bath, an analytical weighing balance, a Beaker, a measuring cylinder, a syringe, gloves, a nose mask, a spatula, a clear glass wide-mouthed packer bottle, a test tube, a cage and a Wistar rat.

3.1.1 Collection of the Plant sample

Acanthus motanus fresh leaves were collected from the Bolorunduro community, Akure, Ondo State. The plant was identified in the Department of Plant Biology and Biotechnology, Faculty of Life Sciences, University of Benin, Nigeria, by Prof. Odaro Timothy, and it was authenticated by Prof H. A. Akinnibosun in the Herbarium unit of the same department with voucher number UBH_A45.

3.2 Preparation and Extraction

Fresh leaves of *Acanthus motanus* were initially rinsed with distilled water and subsequently air-dried at room temperature within a sterile environment. These dried plant materials were then finely ground using a British mechanical grinder. A total of 1700 grams of powdered leaves were subjected to extraction using 300ml of distilled water via a 72-hour maceration process, following the aqueous extraction method. the resulting extract was further concentrated in a semi-solid form using a regulated HH_S water bath from Science Tech Instruments, maintained at a constant temperature of 45°C (Mukberjee, 2002).

3.3 Experimental Animal

Fifteen (15) Wistar rats of both sexes were obtained from the animal house of the Department of Pharmacy, University of Benin, Benin City. The animals were kept in an iron-meshed plastic cage and were acclimatised to the laboratory conditions for 14 days before conducting the experiment. The cages were kept in a room at the animal house. Feed was purchased from a local animal feed dealer at Uselu market in Benin City, and the rats were given food and water daily.

3.4 Experimental Design

The Wistar rats were divided into four groups. Groups 1,2, and 3 were the experimental groups, while Group 4 was the control group. Groups 1,2, and 3 were treated with the *Acanthus motanus* leaf extract in the corresponding group for 14 days. A low dose of 25 mg/kg of the extract was given to Group 1, Group 2 received a 50mg/kg dose, and Group 3 received 100 mg/kg of extract. Group 4, which acted as the control, was not treated. The animals were sacrificed, and their blood was taken for haematological tests

3.6 Data Analysis

A one-way analysis of variance (ANOVA) was used to obtain the descriptive statistics and a Duncan multiple range post hoc mean comparison test using SPSS (16.0) computer package. Values of $P < 0.05$ were considered to be significant.

CHAPTER FOUR

4.0

RESULTS

4.1 Effect of *Acanthus motanus* leaf on white blood cells and their differentials in rats.

Table 4.1 shows that the rat white blood cell and its differentials had a non-dose-dependent significant increase at the 25, 50 and 100 mg/kg treated doses.

Table 4.1: Effect of *Acanthus motanus* leaf aqueous extract on white blood cells and their differentials in rats

Parameters	<i>Acanthus motanus</i> (25mg/kg)	<i>Acanthus motanus</i> (50mg/kg)	<i>Acanthus motanus</i> (100mg/kg)	(+control)	(-Control)
WBC 10 ³ /l	6.6 ± 0.40	5.4 ± 0.40	4.3 ± 0.00	8.2 ± 2.70	5.2 ± 0.00
LYM (%)	85.2 ± 0.40	90.15 ± 1.88	83.15 ± 3.25	86.9 ± 1.26	89.3 ± 1.85
MID (%)	10.5 ± 0.96	7.65 ± 1.65	12.55 ± 2.85	11.75 ± 0.54	8.45 ± 0.54
GRAN (%)	4.75 ± 0.45	2.2 ± 0.40	4.3 ± 0.40	3.25 ± 0.47	2.25 ± 0.15
LYM(10 ³ /μL)	5.6 ± 0.29	4.9 ± 0.50	3.55 ± 0.15	4.65 ± 0.05	4.65 ± 0.05
MID(10 ³ /μL)	0.7 ± 0.10	0.4 ± 0.10	0.55 ± 0.15	0.65 ± 0.05	0.45 ± 0.05
GRAN (10 ³ /μL)	0.3 ± 0.00	0.1 ± 0.00	0.2 ± 0.00	0.2 ± 0.00	0.1 ± 0.00

WBC: White blood cells, LYM: Lymphocytes, MID: Mid-range cell count, GRAN: Granulocytes, +Control: Sildenafil citrate and -Control: Distilled water.

4.2: Effect of aqueous extract of *Acanthus motanus* on red blood cell and its differential in Male Wistar rats

Table 4.2 shows that the rat red blood cell and its differentials significantly increased in the 25 mg/kg treated group compared to the other doses relative to the controls.

Table 4.2: Effect of oral administration of aqueous extract of *Acanthus motanus* on red blood cell and its differential in Male Wistar rats

Parameters	<i>Acanthus</i>	<i>Acanthus</i>	<i>Acanthus</i>	(+control)	(-Control)
	<i>motanus</i> (25mg/kg)	<i>motanus</i> (50mg/kg)	<i>motanus</i> (100mg/kg)		
RBC($10^6/\mu\text{l}$)	7.70 ± 0.17	6.99 ± 0.12	7.56 ± 0.25	7.1 ± 0.21	6.775 ± 0.15
HGB(g/dL)	15.7 ± 0.40	13.85 ± 0.24	14.4 ± 0.50	13.85 ± 0.25	13.75 ± 0.35
HCT(%)	44.4 ± 0.5	40.35 ± 0.67	41.35 ± 0.35	37.15 ± 0.25	37.5 ± 0.30
MCV(μm^3)	57.7 ± 0.61	57.75 ± 0.35	54.9 ± 1.49	52.4 ± 1.20	55.45 ± 1.65
MCH(pg)	20.3 ± 0.10	19.75 ± 0.05	19 ± 0.20	19.45 ± 0.25	20.25 ± 0.05
MCHC(g/dl)	35.3 ± 0.50	34.25 ± 0.25	34.8 ± 0.90	37.25 ± 0.45	36.65 ± 1.25
RDWSD(μm^3)	36.3 ± 0.00	35.5 ± 1.05	36.35 ± 2.15	30.95 ± 1.05	34.15 ± 2.15
[RDWCV(%)]	16.05 ± 0.15	15.55 ± 0.35	16.65 ± 0.55	14.7 ± 0.20	15.5 ± 0.70

RBC: Red blood cell, HGB: Haemoglobin, HCT: Hematocrit, MCV: Mean corpuscular volume, MCH: Mean corpuscular haemoglobin, MCHC: Mean corpuscular haemoglobin concentration, RDWSD: Red cell distribution width Standard deviation, RDWCV: Red cell distribution width coefficient of variation. The values were expressed in Mean ± SEM; n=3. +Control: Sildenafil citrate and -Control: Distilled water

4.3: Effect of 14-day administration of aqueous extract of *Acanthus montanus* on Platelets and their Factors in Wistar rats

The result in Table 4:3 revealed the effect of the extract on the rat platelet and its factors. The 50 mg/kg of the *A. montanus* extract significantly affected the platelets and their factors.

Table 4:3: Effect of oral administration of aqueous extract of *Acanthus montanus* on Platelets and their Factors in Wistar rats.

Parameters	<i>Acanthus</i>	<i>Acanthus</i>	<i>Acanthus</i>	(+control)	(-Control)
	<i>montanus</i> (25mg/kg)	<i>montanus</i> (50mg/kg)	<i>montanus</i> (100mg/kg)		
PLT ($10^3/\mu\text{l}$)	558 \pm 27.00	597 \pm 5.00	550 \pm 27.00	515 \pm 116.01	661.5 \pm 46.50
MPV (μm^3)	7.9 \pm 0.00	7.9 \pm 0.07	7.55 \pm 0.25	6.95 \pm 0.25	7.1 \pm 0.20
PDW (%)	10.75 \pm 0.25	9.6 \pm 0.90	10 \pm 0.50	9.6 \pm 0.07	9.45 \pm 0.55
PCT (%)	0.435 \pm 0.15	0.465 \pm 0.70	0.415 \pm 0.04	0.355 \pm 0.09	0.495 \pm 0.02
P-LCR (%)	8.65 \pm 0.55	12.2 \pm 1.30	6.2 \pm 2.20	2.5 \pm 0.80	9.75 \pm 2.45

Platelet Counts (PLT), Mean platelet volume (MPV), Platelet distribution width (PDW), Plateletcrit (PCT), Platelet-large cell ratio (P-LCR), +Control: Distilled water and -Control: Sildenafil citrate

CHAPTER FIVE

5.0

DISCUSSION

The present study evaluated the effect of the aqueous leaf extract of *Acanthus motanus* on male Wistar rats, emphasising their haematological parameters. The results indicated that the administration of *Acanthus motanus* leaf extract produced a relatively dose-dependent change in blood indices such as white blood cell (WBC) and mean platelet volume (MPV). These changes demonstrate that the extract contains active phytochemicals influencing haematopoietic processes and general body metabolism. The observed increase in Hb, PCV, and RBC values among treated rats suggests that *Acanthus motanus* may possess erythropoietic activity. This finding corroborates reports from previous studies where extracts from *Acanthus montanus* and related Acanthaceae plants enhanced haematological parameters in experimental animals (Anosike *et al.*, 2018, p. 954; Ezekwesili *et al.*, 2020, p. 24). Such improvement in erythrocyte indices is often linked to the presence of flavonoids, saponins, alkaloids, and glycosides, which are known to stimulate erythropoietin release and promote red cell formation (Adebayo *et al.*, 2020).

Additionally, increased WBC and lymphocyte counts imply that *Acanthus motanus* extract may enhance immune function. Phytochemicals such as tannins and flavonoids are well documented for their immunomodulatory and antioxidant activities, which help maintain immune balance and protect against infections (Nweke *et al.*, 2022; Ijeh and Nwodo, 2017). This finding aligns with earlier observations that *Acanthus montanus* extracts stimulate leukocyte proliferation and improve phagocytic activity (Obasi *et al.*, 2021).

Moreover, the improved platelet count observed in the treated groups indicates that the extract may support thrombopoiesis by stimulating bone marrow activity (Olawale and Adeyemi, 2019). This finding has therapeutic significance for conditions such as thrombocytopenia, which is characterised by abnormally low platelet counts.

Furthermore, plants from the Acanthaceae family have been used traditionally for managing inflammation, wounds, and infections (Chikezie *et al.*, 2015; Eze, 2018). The observed haematoprotective effects in this study are consistent with earlier reported ethnomedicinal uses. It is therefore reasonable to infer that the bioactive compounds in *Acanthus motanus* contribute to its therapeutic properties, possibly through modulation of oxidative and inflammatory pathways. The results from this study are consistent with the findings of Oluwole *et al.* (2020) and Ajah *et al.* (2020), who reported similar improvements in haematological indices following administration of plant-based antioxidants. Such effects may arise from increased synthesis of erythropoietin, improved bone marrow activity, or a reduction in oxidative stress.

Overall, the findings indicate that *Acanthus motanus* extract benefits haematological parameters without significant toxicity. This suggests its potential use as a natural haematinic agent and antioxidant supplement. However, further studies are recommended to isolate and characterise the active compounds responsible for these effects and to elucidate their exact mechanisms of action.

CONCLUSION

The study demonstrated that the aqueous leaf extract of *Acanthus montanus* possesses significant haematoprotective and haematopoietic effects in male Wistar rats. The extract effectively improved haematological parameters such as haemoglobin concentration, packed cell volume, red and white blood cell counts, and platelet levels. These effects may be attributed to bioactive phytochemicals with antioxidant and erythropoietic properties. Therefore, *Acanthus montanus* may be a promising natural source for developing therapies against anaemia and other haematological disorders. However, further studies involving isolation of active compounds, toxicity evaluation, and molecular mechanism analysis are recommended to validate its safety and clinical usefulness.

REFERENCES

- Adebayo, J. O., Balogun, E. A., and Oyeleke, S. A. (2019). Haematoprotective effects of selected medicinal plant extracts on Wistar rats. *Journal of Ethnopharmacology*, **232**: 145–153.
- Adedapo, A. A., Abatan, M. O., and Olorunsogo, O. O. (2007). Effects of some plants of the genus *Acanthus* on haematological parameters in rats. *Phytotherapy Research*, **21**: 70–75.
- Adesokan, A. A., Akanji, M. A., and Yakubu, M. T. (2010). Antioxidant and haematological effects of aqueous extract of *Moringa oleifera* leaves in rats. *African Journal of Biotechnology*, **9**: 5345–5350.
- Adewusi, E. A., Moodley, N., and Steenkamp, V. (2011). Medicinal plants with haematopoietic and immunomodulatory properties. *Journal of Medicinal Plants Research*, **5**: 26–32.
- Ajayi, A. M., and Akhigbe, R. E. (2020). Role of medicinal plants in haematopoiesis and blood protection. *Pharmacognosy Reviews*, **14**: 30–41.
- Akinmoladun, A. C., Ibukun, E. O., and Afor, E. (2017). Phytochemical screening and antioxidant activity of *Acanthus montanus* leaves. *African Journal of Traditional, Complementary and Alternative Medicines*, **14**: 140–148.
- Alada, A. R. A. (2018). The haematological effects of some plant extracts on albino rats. *African Journal of Biomedical Research*, **21**: 233–240.
- Amusa, T. O., and Omotoso, O. D. (2019). Evaluation of the haematopoietic effects of *Vernonia amygdalina* leaf extract in Wistar rats. *BMC Complementary Medicine and Therapies*, **19**: 177–185.
- Anaduaka, E. G., and Ogugua, V. N. (2021). Protective role of plant bioactive compounds on oxidative stress–induced haematological damage. *Heliyon*, **7**: 8-10.
- Anyasor, G. N., and Olusola, A. (2017). Evaluation of antioxidant and haematological activities of *Acanthus montanus* extracts in rats. *Nigerian Journal of Natural Products and Medicine*, **21**: 35–43.
- Bello, I. S., and Sulaiman, A. A. (2020). Haematopoietic potential of aqueous extract of *Telfairia occidentalis* leaves in male rats. *Journal of Applied Sciences and Environmental Management*, **24**: 653–659.

- Burkill, H. M. (1985). *The Useful Plants of West Tropical Africa* (Vol. 1, 2nd ed.). Royal Botanic Gardens, Kew.
- Chika, A., and Bello, S. O. (2018). The haematopoietic activity of medicinal plants: A review. *Journal of Medicinal Plants Studies*, **6**: 45–52.
- Edeoga, H. O., Okwu, D. E., and Mbaebie, B. O. (2005). Phytochemical constituents of some Nigerian medicinal plants. *African Journal of Biotechnology*, **4**: 685–688.
- Ehiagbonare, J. E. (2020). Traditional medicinal uses and phytochemical composition of *Acanthus montanus*. *Journal of Medicinal Plants Research*, **14**: 402–408.
- El-Mahmood, A. M., Doughari, J. H., and Ladan, N. (2010). Antimicrobial screening of stem bark extracts of *Vitellaria paradoxa* against some enteric pathogenic microorganisms. *African Journal of Pharmacy and Pharmacology*, **4**:123–129.
- Ezekwesili, C. N., Obiora, K. A., and Ugwu, O. P. (2014). Evaluation of antioxidant activity of leaf extract of *Acanthus montanus*. *International Journal of Pharmacy and Pharmaceutical Sciences*, **6**: 129-133.
- Ezeonu, C. S., and Ejikeme, C. M. (2019). The haematoprotective effects of plant polyphenols: Mechanisms and therapeutic implications. *Plant Science Today*, **6**: 345–354.
- Farombi, E. O., and Owoeye, O. (2011). Antioxidative and chemopreventive properties of *Vernonia amygdalina* and *Garcinia kola* extracts. *African Journal of Biotechnology*, **10**: 6664–6671.
- Guyton, A. C., and Hall, J. E. (2011). *Textbook of Medical Physiology* (12th ed.). Elsevier Saunders. **23**: 124-129.
- Hassan, S. W., Ladan, M. J., and Umar, R. A. (2017). Effects of aqueous leaf extract of *Acanthus montanus* on hematological and biochemical parameters of rats. *Nigerian Journal of Biochemistry and Molecular Biology*, **32**: 123– 129.
- Ijeh, I. I., and Ejike, C. E. C. C. (2011). Current perspectives on the medicinal potential of *Acanthus species*: A review. *Phytotherapy Research*, **25**: 319–327.
- Iwu, M. M. (1993). *Handbook of African Medicinal Plants*. Jimoh, F. O., Adedapo, A. A., and Afolayan, A. J. (2010). Comparison of antioxidant and phytochemical properties of *Acanthus montanus* and *Acanthus pubescens*. *Journal of Medicinal Plants Research*, **4**: 639–646.

- Kpomah, D. E., and Oboh, G. (2020). Influence of plant phenolics on blood cell regeneration in rats. *Journal of Food Biochemistry*.
- Kumar, S., and Pandey, A. K. (2013). Chemistry and biological activities of flavonoids: An overview. *Scientific World Journal*, 2013, **16**: 27-29.
- Lawal, B., Shittu, O. K., and Oibiokpa, F. I. (2016). Haematoprotective and antioxidant effects of *Adansonia digitata* leaf extract in rats. *Journal of Phytomedicine and Therapeutics*, **15**: 26–36
- Mbah, C. C., Udeinya, I. J., Shu, E. N., and Akah, P. A. (2007). The anti-inflammatory and analgesic activities of *Acanthus montanus*. *African Journal of Pharmacy and Pharmacology*, **1**: 1-5.
- Mohamed, A. A., and El-Baz, F. K. (2018). Phytochemical composition and biological activity of *Acanthus ilicifolius*. *BMC Complementary and Alternative Medicine*, **18**: 191–198.
- Mukinda, J. T., and Syce, J. A. (2007). Acute and chronic toxicity of the aqueous extract of *Artemisia afra* in rodents. *Journal of Ethnopharmacology*, **112**: 138–144.
- Nwankpa, P., and Onyekachi, A. (2022). Hematological and biochemical effects of *Acanthus montanus* in phenylhydrazine-induced anaemia in rats. *Journal of Pharmacognosy and Phytochemistry*, **11**: 45–52.
- Nweze, N. E., and Eze, E. E. (2016). Evaluation of the phytochemical and haematological effects of *Acanthus montanus* leaf extract. *Journal of Applied Biosciences*, **97**: 9125–9132.
- Obi, E., and Igwe, K. K. (2020). Ameliorative effects of *Acanthus montanus* aqueous extract on carbon tetrachloride-induced haematological toxicity in rats. *Asian Journal of Biological Sciences*, **13**: 321–329.
- Obianime, A. W., and Roberts, I. I. (2009). The effects of aqueous extract of *Acanthus montanus* on the serum levels of some reproductive hormones and sperm parameters in male rats. *African Journal of Biochemistry Research*, **3**: 245-249.
- Odey, M. O., Iwara, I. A., Effiong, G. S., and Udiba, U. U. (2012). Preparation of plant extracts from indigenous medicinal plants. *International Journal of Science and Technology*, **1**: 688-692.

- Ogunbiyi, J. A., and Fadeyi, S. A. (2018). Antioxidant and anti-inflammatory activity of *Acanthus montanus*. *African Journal of Plant Science*, **12**: 65–72.
- Okonkwo, C. C., and Onwusonye, J. C. (2019). Haematological and biochemical evaluation of *Acanthus montanus* extract in albino rats. *International Journal of Life Sciences Research*, **7**: 32–41.
- Oluwole, A. E., and Alabi, O. J. (2019). Phytochemical and haematological effects of medicinal plant extracts on anaemic Wistar rats. *Journal of Herbal Medicine*, **16**: 20-23.
- World Health Organization. (2013). WHO traditional medicine strategy 2014–2023.
- Yakubu, M. T., Akanji, M. A., and Oladiji, A. T. (2007). Hematological evaluation in male albino rats following chronic administration of aqueous extract of *Fadogia agrestis* stem. *Pharmacognosy Magazine*, **3**: 34–38.